This article was downloaded by: On: 22 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



To cite this Article Huang, S. -X., Zhou, Y., Nie, Q. -J., Ding, L. -S. and Peng, S. -L.(2006) 'Two new iridoid glucosides from *Picrorhiza scrophulariiflora*', Journal of Asian Natural Products Research, 8: 3, 259 — 263 To link to this Article: DOI: 10.1080/10286020500034543 URL: http://dx.doi.org/10.1080/10286020500034543

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Two new iridoid glucosides from Picrorhiza scrophulariiflora

Journal of Asian Natural Products Research, Vol. 8, No. 3, April-May 2006, 259-263

S.-X. HUANG[†], Y. ZHOU[†], Q.-J. NIE[‡], L.-S. DING[†] and S.-L. PENG^{†*}

*Chengdu Institute of Biology, Chinese Academy of Sciences, Chengdu 610041, China *Sichuan Drug Administration, Division of Drug Licence, Chengdu 610015, China

(Received 17 August 2004; revised 9 November 2004; in final form 19 November 2004)

Two new iridoid glucosides with 3,4-dihydrocatalpol skeleton, piscrosides A (1) and B (2) together with nine known iridoid glucosides and three known cucurbitacin glucosides, were isolated from the stems of *Picrorhiza scrophulariiflora*. Their structures were established by MS, ¹H NMR, ¹³C NMR and 2D NMR methods (including HSQC, HMBC and NOESY experiments).

Keywords: Picrorhiza scrophulariiflora; Scrophulariaceae; Iridoid glucoside; Piscrosides A and B

1. Introduction

The genus Picrorhiza is widely distributed in China, Nepal, and India, and contains two species, P. scrophulariiflora Pennell and P. kurrooa Royle ex Benth (Scrophulariaceae). The plant, P. kurrooa, was a traditional medicine used in India for treatment of jaundice, indigestion, common fever, acute viral hepatitis, and bronchial asthma [1]. Previous pharmacological studies for this species have revealed hepatoprotective [2], immunostimulative [3], anti-asthma [4], neuritogenic [5], anti-inflammatory [6] and scavenging free radical activities [7]. The other species, P. scrophulariiflora, is a traditional Chinese medicine, which also has hepatoprotective, choleretic, immunostimulative and antiinflammatory activities [8,9]. To date there are several chemical constituents reported from P. scrophulariiflora, except for our work dealing with some new compounds including three phenyl glycosides and two phenylethyl glycosides [10]. During our systematic chemical investigation, two new iridoid glucosides, piscrosides A (1) and B (2), were isolated from the stems of *P. scrophulariiflora*, together with nine known iridoid glucosides, picroside I (3), picroside II (4), picroside III (5), picroside IV (6), 6-O-trans-feruloylcatalpol (7), minecoside (8), verminoside (9), catalposide (10), and aucubin (11) [11,12] (figure 1). Additionally, the stems of the plant afforded only three cucurbitacin glucosides, 2β glucopyranosyloxy-3,16,20,22-tetrahydroxy-9-methyl-19-norlanosta-5,24-diene (12) [13], 2β-glucopyranosyloxy-3,16,20,22-tetrahydroxy-19-norlanosta-5,23-diene-22-ol (13) [14],

^{*}Corresponding author. E-mail: pengsl@cib.ac.cn

S.-X. Huang et al.



and 25-aceloxy-2 β -glucopyranosyloxy-3,16,20-trihydroxy-19-norlanosta-5,23-diene-22-ol (14) [14]. The known compounds were identified by comparisons of spectroscopic data with those of reported ones.

2. Results and discussion

Piscroside A (1) was obtained as an amorphous powder. The positive ESI-MS showed a pseudomolecular ion $[M + Na]^+$ at m/z 537 and the negative ESI-MS exhibited the ions $[M - H]^{-}$ at m/z 513. All these data were compatible with the molecular formula $C_{23}H_{30}O_{13}$, which was also confirmed by HRFAB-MS at m/z [M - H]⁻ 513.1571. The IR spectrum showed absorption bands at 3419 (br OH), 1702 (C=O), and 1605, 1516, 1430 (aromatic ring) cm⁻¹. In the ¹H NMR spectrum, **1** showed three aromatic protons [δ 7.44 (d, J = 1.8 Hz), 7.46 (dd, J = 1.8, 8.4 Hz), 6.87 (d, J = 8.4 Hz)], one phenolic hydroxyl proton (δ 9.95, s) and a methoxy protons (δ 3.81, s), due to the vanilloyl ester part of the molecule. Besides these, 1 resembled those of catalpol, except for the absence of the olefinic signals attributable to H-3 and H-4. The signals corresponding to this double bond were also absent from the ¹³C NMR spectrum of 1, where two signals at δ 61.7 and 23.1 suggest that it has the structure of 3,4-dihydrocatalpol unit. In the HMBC spectrum, the correlations of H-5 $(\delta 1.80)$ and H-1 $(\delta 4.46)$ with C-3 $(\delta 61.7)$, H-6 $(\delta 3.77)$, H-5 $(\delta 1.80)$ and H-9 $(\delta 2.04)$ with C-4 (δ 23.1) were observed. Additionally, the clear correlations, H₂-6' (δ 4.32) with C=O (δ 166.1), enabled us to assign the position of the vanilloyl group to C-6' of glucosyl group. The relative configurations of C-1, C-5, C-6, C-7, C-8, and C-9 were deduced from the results of the NOESY spectrum (figure 2). In the ESI-MSⁿ analysis, MS¹ gave a pseudomolecular ion at m/z 513 [M – H]⁻, MS² gave a fragment ion at m/z $311 [M - H - 202]^{-}$, and MS³ gave the fragment ion at $m/z 167 [M - H - 202 - 144]^{-}$, which

260



Figure 2. The key correlations of HMBC and NOESY of 1.

corresponds to the sequential loss of an iridoid unit and a glucosyl group. Thus, **1** was unambiguously identified as 6'-O-vanilloyl-3,4-dihydrocatalpol, named piscroside A.

Piscroside B (2) was also obtained as amorphous powder. Based on the HRFAB-MS, the pseudomolecular ion peak at m/z 509.1615 [M – H], the molecular formula of 2 was determined as C₂₄H₃₀O₁₂. Comparison of the ¹H NMR and ¹³C NMR data of 2 with those of 1 indicated the existence of the same skeleton of 3,4-dihydrocatalpol. The ¹H NMR spectrum of 2 showed the presence of a *p*-coumaroyl group, which was confirmed by the four aromatic proton signals between δ 6.79 and 7.57 for an AA'BB' systems [δ 7.57 (2H, *d*, *J* = 8.5 Hz), 6.79 (2H, *d*, *J* = 8.5 Hz)], a *trans*-double bond proton [δ 7.56 (1H, *d*, *J* = 15.9 Hz)], 6.40 (1H, *d*, *J* = 15.9 Hz)], and an aromatic hydroxyl proton [δ 10.02 (1H, *s*)]. All further essential spectral data of 2 were in full agreement with those of 1. Thus, compound 2 was determined to be 6'-O-p-coumaroyl-3,4-dihydrocatalpol, named piscroside B.

Only two iridoid glucosides containing a 3,4-dihydrocatalpol unit have been reported so far since the isolation of the first one in 1979 [15,16]. In addition, the previous and this investigation showed that the two species, *P. scrophulariiflora* and *P. kurrooa*, are similar in the chemical constituents, both mainly containing iridoid glucosides. However, the difference is that cucurbitacin glucosides are far more abundant in the species of *P. kurrooa* than that of *P. scrophulariiflora*.

3. Experimental

3.1 General experimental procedures

IR spectra were recorded on Perkin–Elmer Spectrum One FT-IR spectrometer. NMR spectra were recorded on Bruker AV-600 spectrometer using tetramethylsilane (TMS) as an internal standard. Optical rotations were measured with PE-341 polarimeter. VG AutoSpec-3000 spectrometer was used to record HRFAB-MS and Finnigan LCQ^{DECA} to record ESI-MS and tandem MS spectra. Sephadex LH-20 (Pharmacia), silica gel (200–300 mesh, Qingdao Marine Chemical Group Co.), Lobar LiChroprep RP-18 (40–63 μ m, Merck), Lobar LiChroprep Si-60 (40–63 μ m, Merck) were used for column chromatography.

S.-X. Huang et al.

3.2 Plant material

The dried stems of *Picrorhiza scrophulariiflora* were collected in Tibet of China, and identified by Prof Zuo-Cheng Zhao. A voucher specimen is deposited in the Herbarium of Chengdu Institute of Biology, Chinese Academy of Sciences.

3.3 Extraction and isolation

The dried stems of *P. scrophulariiflora* (6 kg) were extracted with EtOH at 60°C for 3 times, each for 8 h. The concentrated extract was suspended in H₂O and partitioned successively with petroleum ether, chloroform and EtOAc/EtOH (9:1) mixture. The EtOAc/EtOH fraction (50 g) was subjected to column chromatography over silica gel eluted with CHCl₃/CH₃OH (40:1–2:1) to obtain six fractions. Fraction I (6 g) was separated by silica gel column chromatography with CHCl₃/CH₃OH gradient system (20:1–10:1) to give compound **14**. Fraction II (10 g) was chromatographed on ODS column eluting with MeOH/H₂O (1:1) to afford two crude compounds, which were purified by silica gel column chromatography eluting with CHCl₃/MeOH to give **3** (2.1 g) and **12** (100 mg), respectively. Fraction III (2 g) was chromatographed on a Lobar LiChroprep Si-60 column using CHCl₃/CH₃OH (10:1) to

Position	1		2	
	¹ H*	$^{13}C^{\dagger}$	$^{I}H^{*}$	$^{13}C^{\dagger}$
1	4.46 d (8.7)	96.5 d	4.51 d (8.7)	96.6 d
3α	3.74 m	61.7 t	3.75 m	61.7 t
3β	3.19 [‡]		3.21 [‡]	
4α	1.44 d (13.5)	23.1 t	1.45 d (13.5)	23.1 t
4β	1.58 m		1.59 m	
5	1.80 dd (13.5, 8.1)	37.2 d	1.83 dd (13.5, 8.1)	37.2 d
6	3.77 dd (9.0, 6.6)	71.6 d	3.79 dd (9.0, 6.6)	71.6 d
7	3.18 d (9.0)	60.9 d	3.19 d (9.0)	61.0 d
8		65.2 s		65.3 s
9	2.04 t (8.0)	42.4 d	2.06 t (8.0)	42.5 d
10a	3.41 dd (13.2, 7.3)	59.7 t	3.46 dd (13.2, 7.3)	59.7 t
10b	3.81 [‡]		3.84 dd (13.2, 7.3)	
1'	4.55 d (7.8)	98.3 d	4.54 d (7.8)	98.4 d
2'	3.02dd (7.8, 5.0)	74.0 d	3.03 dd (7.8, 5.0)	73.9 d
3'	3.21 [‡]	76.9 d	3.22 [‡]	76.9 d
4′	3.20 [‡]	70.9 d	3.23 [‡]	70.7 d
5'	3.47 m	74.6 d	3.40 m	74.5 d
6′a	4.44 br d (12.0)	64.1 t	4.34 br d (12.0)	65.3 t
6′b	4.32 dd (12.0, 6.3)		4.25 dd (12.0, 6.0)	
1″		121.3 s		125.7 s
2″	7.44 d (1.8)	113.3 d	7.57 d (8.5)	131.0 d
3″		148.0 s	6.79 d (8.5)	116.5 d
4″		152.2 s		160.5 s
5″	6.87 d (8.4)	115.8 d	6.79 d (8.5)	116.5 d
6"	7.46 dd (8.4, 1.8)	124.1 d	7.57 d (8.5)	131.0 d
OCH ₂	3.81 s	56.3 a		10110 0
CO	2.01.0	166.1 s		167.2 s
α			6.40 d (15.9)	114.7 d
ß			7.56 d (15.9)	145.5 d

Table 1. ¹H NMR and ¹³C NMR data for compounds 1 and 2.

* 600 MHz, DMSO- d_6 ; chemical shifts in ppm relative to TMS; coupling constant (J) in Hz

[†]150 MHz, DMSO- d_6 ; multiplicity was established from DEPT data

[‡]Overlapped.

give two fractions; one fraction was purified by repeatedly silica gel to obtain compound **6** (35 mg). The other fraction was submitted to column chromatography (RP-18) with MeOH/H₂O (1:1) to yield compounds **5** (20 mg) and **13** (17 mg), respectively. Compounds **4** (3.5 g), **7** (15 mg), and **8** (150 mg) were obtained from fraction IV (20 g) by a Lobar LiChroprep RP-18 column. Fraction V (5 g) was repeatedly chromatographed on ODS gel (eluted with H₂O \rightarrow 70% MeOH), Lobar LiChroprep RP-18 (eluted with H₂O \rightarrow 60% MeOH), Lobar LiChroprep Si-60 [eluted with CHCl₃ \rightarrow CHCl₃/MeOH (4:1)] and Sephadex LH-20 (eluted with MeOH) to give compounds **1** (15 mg), **2** (5 mg), **9** (14 mg) and **10** (8 mg), respectively. Compound **11** (15 mg) was obtained from the last fraction (4.5 g) by chromatography on silica gel column.

3.4 Identification

Piscroside A (1): White amorphous powder. $[\alpha]_D^{25} - 59.8$ (*c* 0.40, MeOH). IR (KBr): v_{max} 3419, 2925, 1702, 1605, 1516, 1430, 1286, 1221, 1070, 763 cm⁻¹. ESI-MS: *m/z* 513 [M - H], 549 [M + Cl], 537 [M + Na]⁺. HRFAB-MS: *m/z* 513.1571 [M - H] (calcd for C₂₃H₃₀O₁₃, 513.1608). ¹H NMR and ¹³C NMR: see table 1.

Piscroside B (2): White amorphous powder. $[\alpha]_D^{25} - 60.1$ (*c* 0.10, MeOH). IR (KBr): v_{max} 3380, 2918, 1661, 1610, 1580, 1511, 1280, 1241, 1066, 974, 772 cm⁻¹. ESI-MS: *m/z* 509 [M - H], 533 [M + Na]⁺. HRFAB-MS: *m/z* 509.1615 [M - H] (calcd for C₂₄H₃₀O₁₂, 509.1659). ¹H NMR and ¹³C NMR: see table 1.

References

- H. Zhang, Z. Zhang. Handbook of Chinese Traditional Medicine Resources, p. 1149, Science Press, Beijing (1994).
- [2] B.N. Dhawan. Med. Chem. Res., 5, 595 (1995).
- [3] F. Engels, B.F. Renirie, B.A. Hart, R.P. Labadie, F.P. Nijkamp. FEBS Lett., 305, 254 (1992).
- [4] W. Dorsch, H. Stuppner, H. Wagner, M. Gropp, S. Demoulin, J. Ring. Int. Arch. Allergy. Appl. Immunol., 95, 128 (1991).
- [5] P. Li, K. Matsunaga, T. Yamakuni, Y. Ohizumi. Eur. J. Pharmacol., 406, 203 (2000).
- [6] A. Puri, R.P. Saxena, Sumati, P.Y. Guru, D.K. Kulshreshtha, K.C. Saxena, B.N. Dhawan. Planta Med., 58, 528 (1992).
- [7] R. Chander, N.K. Kapoor, B.N. Dhawan. Biochem. Pharm., 44, 180 (1992).
- [8] J. Liu, B.L. Liu, J.Q. Zhang, N. Zhang. Chin. J. New Drugs, 11, 459 (2002).
- [9] H.F. Smit, B.H. Kroes, A.J.J. Van den Berg, D. Van der Wal, E. Van den Worm, C.J. Beukelman, H. Van Dijk, R.P. Labadie. J. Ethnopharmacol., 73, 101 (2000).
- [10] S.X. Huang, X. Liao, Q.J. Nie, L.S. Ding, S.L. Peng. Helv. Chim. Acta, 87, 598 (2004).
- [11] L. El-Naggar, J. Beal. J. Nat. Prod., 43, 649 (1980).
- [12] C.A. Boros, F.R. Stermitz. J. Nat. Prod., 53, 1055 (1990).
- [13] D.Q. Wang, Z.D. He, B.S. Feng, C.R. Yang. Acta Botan Yunnan, 15, 83 (1993).
- [14] H. Stuppner, H. Wagner. Planta Med., 55, 559 (1989).
- [15] R.K. Chaudhuri, O. Sticher. Helv. Chim. Acta, 62, 644 (1979).
- [16] A. Bianco, E. Marini, S.F. Nicoletti, J. Garbarino, M. Piovano, M. Chamy. Phytochemistry, 31, 4203 (1992).